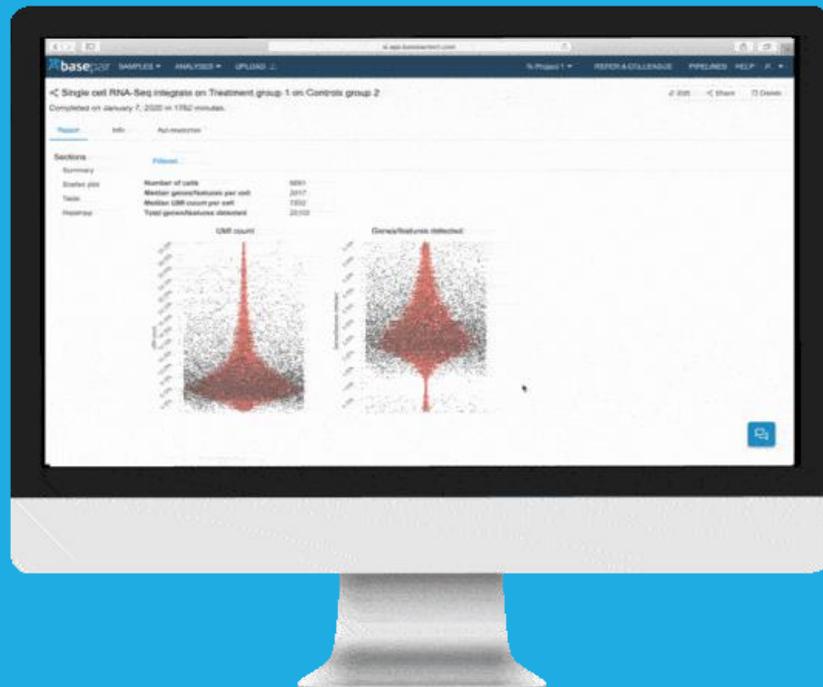


WETもDRYも使える クラウドバイオインフォマティクス環境



薬師寺 秀樹

tech@astride.jp



NGSのデータ解析を行う場合…

NGSのデータ解析を行う場合…

自分でやる

受託会社に依頼する

専用ソフトウェアを使う

NGSのデータ解析を行う場合…

自分でやる

コーディングが…

PCやソフトウェアの設定が…

受託会社に依頼する

前の解析と比較したいのだけど…

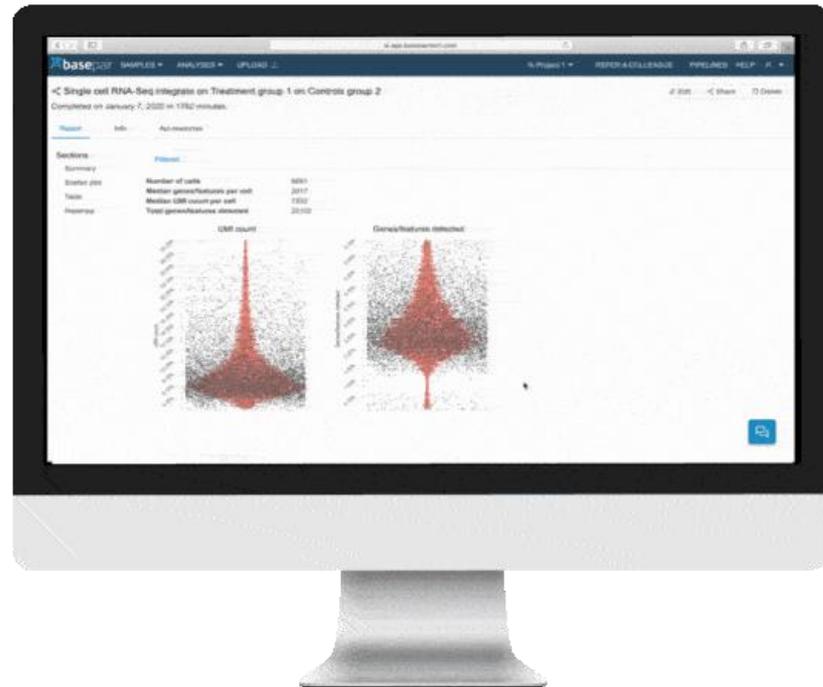
ちょっとした変更なんだけど…

専用ソフトウェアを使う

他の解析ができない…

ライセンスが高い…

 **basepair**



GUIで、QCやアライメント～視覚化まで

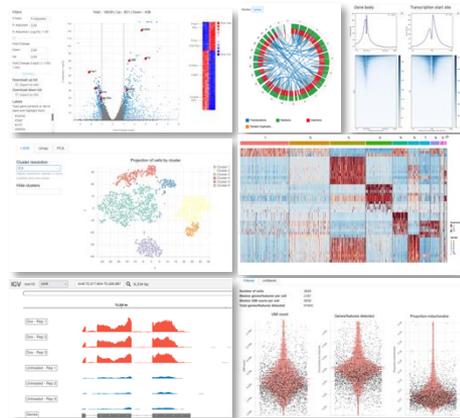
Sequencing



```
>gi|5524211|gb|AAD44166.1| cytochrome  
b [Elephas maximus maximus]  
LCLYTHIGRNIYGSYLSE TWNT GIMILLITMA  
TAFMGYVLPWQMSFWGATVITNLFSAIPYIGT  
NLVEWI VGGFSVDKAT LNRFFAFHFLPFTMVA  
LAGVHLTFLHETGSNNPLGLTSDSDKIPFHPYYT  
IKDFLGLLITLL LLLL LALSPDMLGDPDNHMPAD  
PLNTPLHIKPEWYRLFAYAILRSV PNLGGV LALF  
LSIVLGLMPFLHTSKHRSMM LRP LSQLFWTLT  
MDLL TL TWIGSQPV EYPYTIIGQMASILYFSILA  
FLPIAGX IENY
```

RAW Data
FASTA、BAM etc.

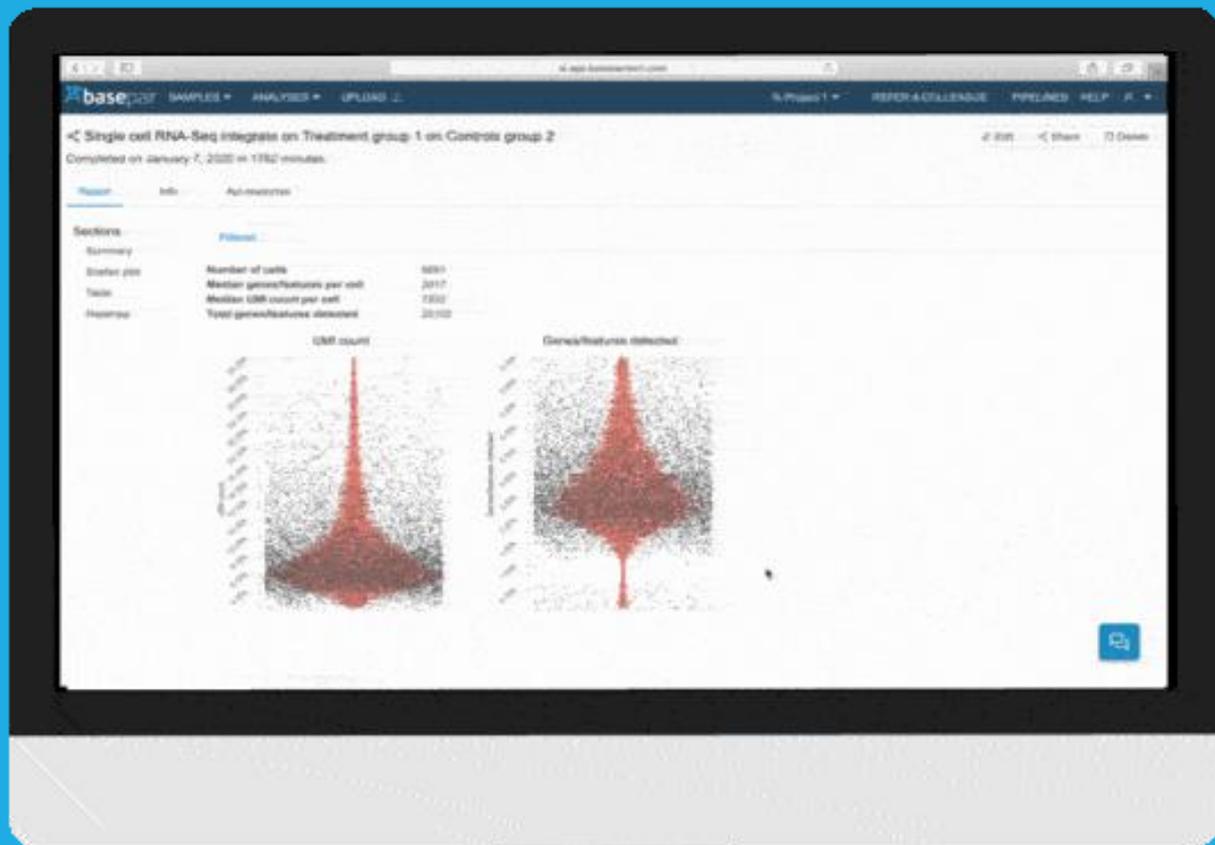
 **basepair**



Analyze

QC, trim, alignment, application related analysis and visualization such as variant call, volcano plot, etc.

マウスで簡単操作 (GUI)



対応済みアプリケーション

✓ ゲノミクス

- ✓ WGS, WES, Panels, CRISPR etc.

✓ トランスクリプトミクス

- ✓ bulk and single cell RNA-Seq

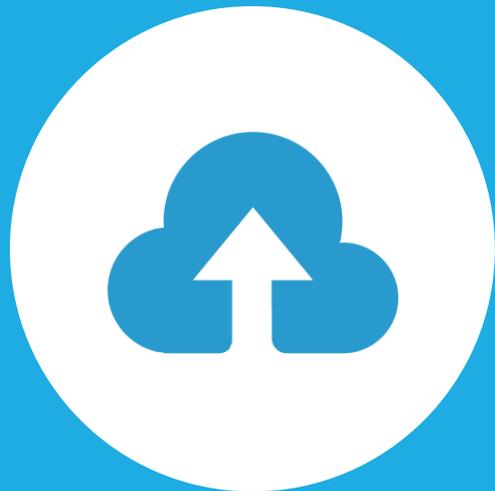
✓ エピゲノミクス

- ✓ ATAC-Seq, ChIP-Seq, CUT&RUN, CUT&TAG etc.

✓ メタジェノミクス

- ✓ 16S, Shotgun etc.

3ステップで解析



1. アップロード

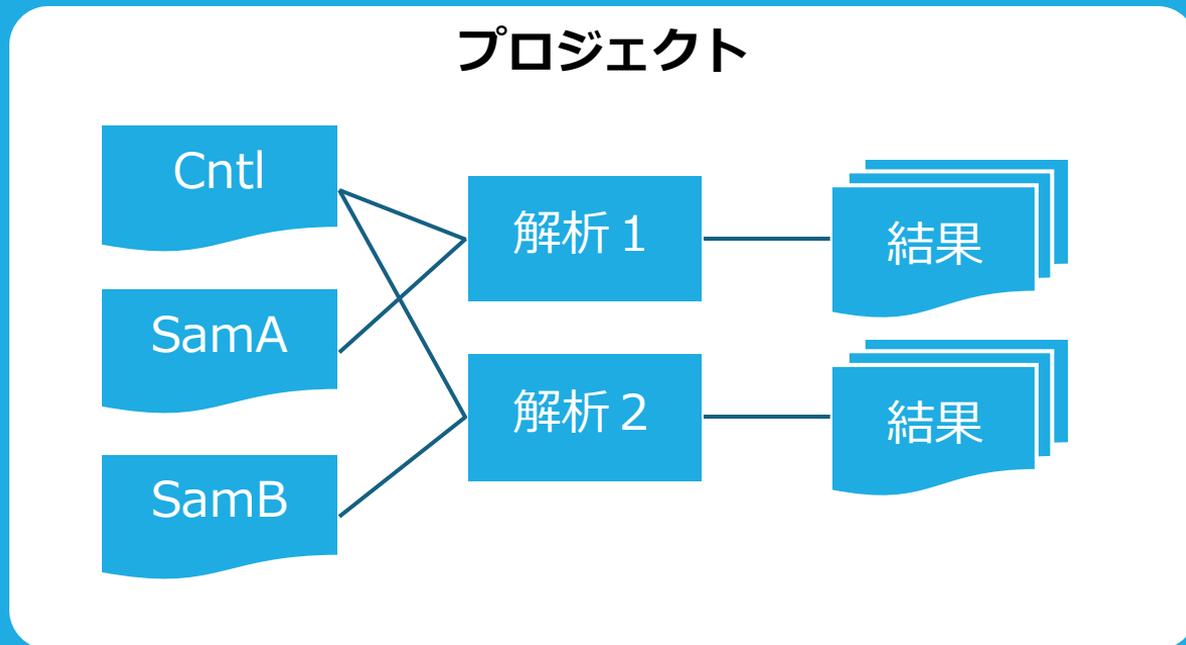


2. 解析



3. 共有

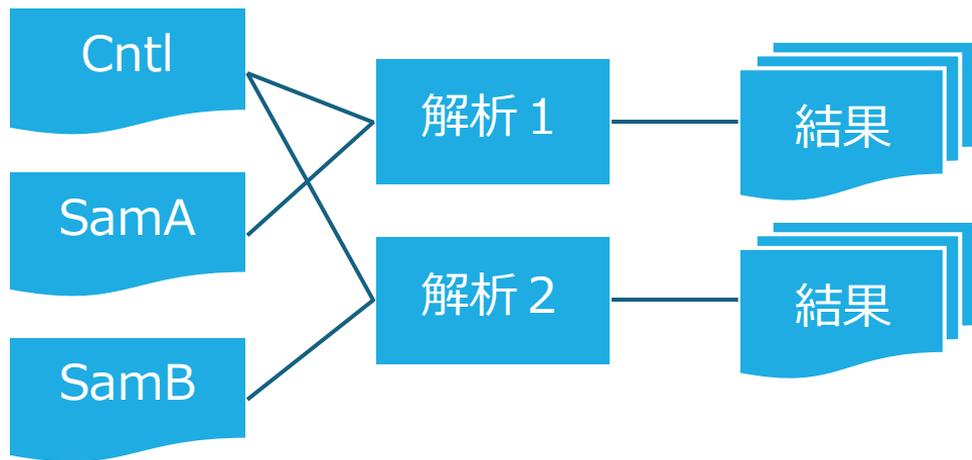
プロジェクト・サンプル・解析



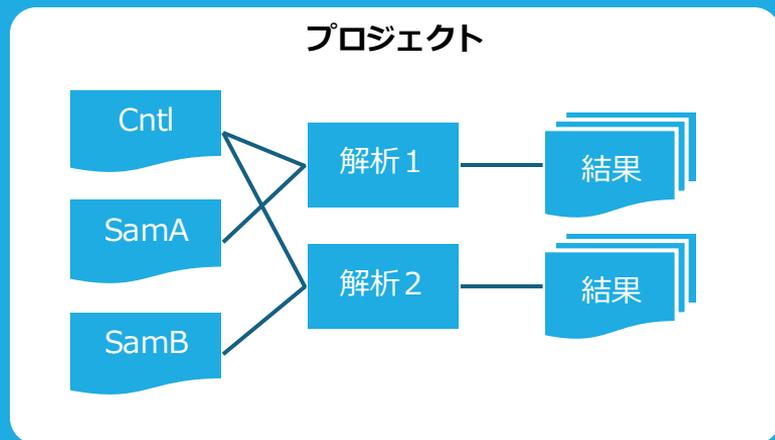
サンプル数により課金・解析数は制限なし

プロジェクト・サンプル・解析

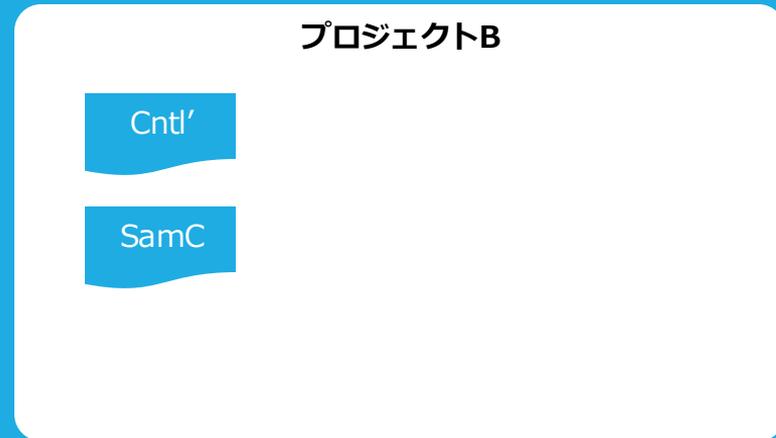
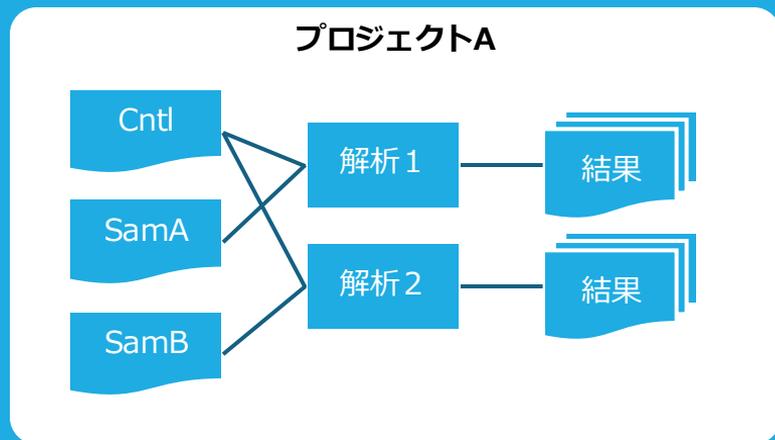
プロジェクト



プロジェクト・サンプル・解析

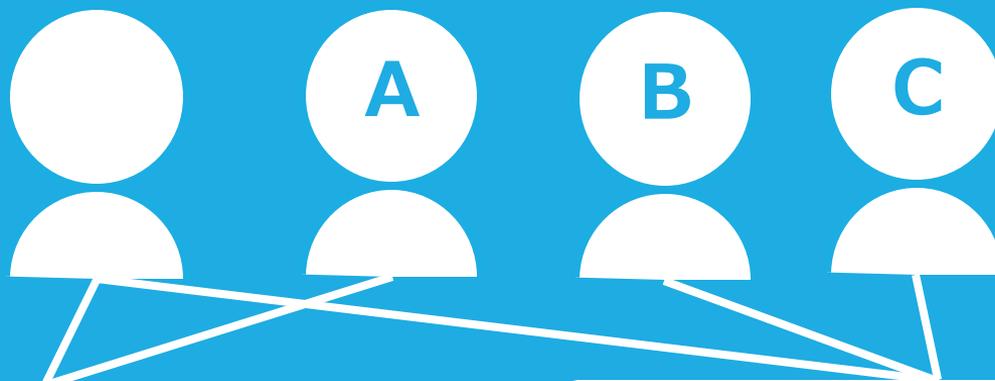


プロジェクト・サンプル・解析

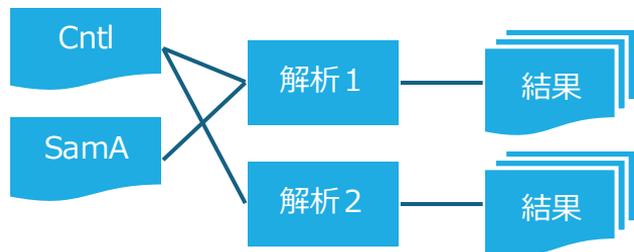


- 複数のプロジェクトを作ることが可能。
- 解析対象は、同じプロジェクト内にある必要がある。
- サンプルのプロジェクト間移動は可能。

共有



プロジェクトA

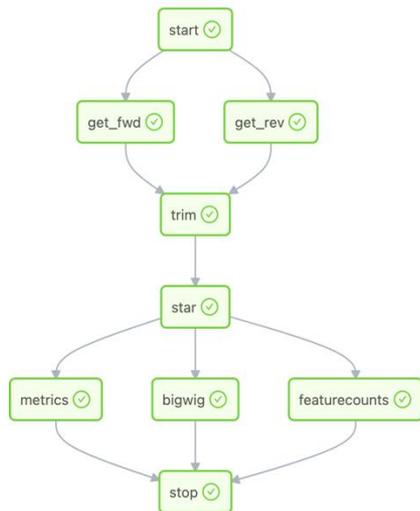


プロジェクトB

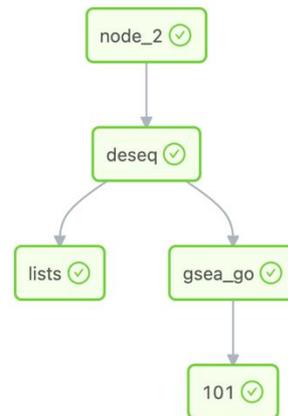


Basepairのパイプライン (Differential Expressionの場合)

Expressio Count (STAR)



Differential Expression (DESeq2)



FastqからDEGまでに必要なステップを接続

サンプルアップロード

New sample



Drag and drop or click here to upload files.
Single and multi-sample upload supported.

① Allowed file formats: .ab1, .bam, .cram, .crai, .csfasta, .csv, .fastq, .fq, .gpr, .gvcf, .qual, .vcf, .sam, .sra, .tsv, .txt, .bz, .bz2, .gz, .zip, .fasta, .fasta.gz

Sample Grouping: Automatic ▼

▼ **Sample name:** 📄 🗑️

▼ Forward ▼ Reverse

📄 04_ATAC2AM_HeLa_rep2_hs_i11_r1.fastq.gz ⬇️ 📄 04_ATAC2AM_HeLa_rep2_hs_i11_r2.fastq.gz ⬇️

▼ **Sample name:** 📄 🗑️

▼ Forward ▼ Reverse

📄 01_ATAC2AM_GM12878_rep1_hs_i5_r1.fastq.gz ⬇️ 📄 01_ATAC2AM_GM12878_rep1_hs_i5_r2.fastq.gz ⬇️

Project: ▼

Tags

Fastqをドラッグ&ドロップ
もしくは、ファイルを選択

FW・RV、サンプルグループ
は自動判定

格納する「プロジェクト」を
選択

Project: Gorota Test1

Tags

Select metadata for all samples

Platform ?

Illumina



Data type ?

ATAC-Seq



Spike in (optional) ?

None



Genome ?

GRCh38



Insert size ?

Pipeline (optional) ?

Ictalurus_punctatus

mouse_GRCm38_M18

Citrus_clementina.v1

Nematostella_vectensis

Gallus_gallus

GRCh38

upload

解析 (Differential Expressionの場合)

New Analysis

* Pipeline Differential Expression (DESeq2, compare 2 groups) ▾

← パイプライン選択 (先読み検索)

Treatment group

* Treatment group name Treatment group 1

* Samples 01-fastq ×

← 処理群 (複数選択可能)

Add samples

Control group

* Controls group name Controls group 2

* Controls 04-fastq ×

← 対照群 (複数選択可能)

Add samples

* Analysis name Differential Expression (DESeq2, compare 2 groups) on Treatment group 1 vs Controls group 2

← 解析名 (任意の名前)

▼ Change default options

> Factors

▼ Deseq

Control group

* Controls group name

* Controls

* Analysis name

▼ Change default options

> Factors

▼ Deseq

Alpha

Features

Hypothesis

Reducedmodel

Minimum expression cutoff

pAdjustMethod

Do not use spike-in for normalization True False

DESeq2のパラメーター

解析開始ボタン



抽出条件

- cell type : cd4
- diseasestatus :
 - type1 diabetes
 - healthy control
- gender : female

使用データ

- GSM1479441
- GSM1479457
- GSM1479463
- GSM1479502
- GSM1479509
- GSM1479523
- GSM1479543

Analysis

Project:

Import data from GEO

[✎ Edit](#) [🔗 Share](#) [🗑 Delete](#) [🔄 Restart](#)

Completed on March 5, 2024 in 120 minutes.

Report

Input/Output

Execution summary

Api response

Sections

Import GEO

Error Output

Import GEO

id	Name	Basepair Id
GSM1479441	lib229	678
GSM1479457	lib245	679
GSM1479463	lib251	680
GSM1479502	lib291	681
GSM1479509	lib298	682
GSM1479523	lib312	685
GSM1479543	lib332	691

Differential Expression (DESeq2, compare 2 groups) on type 1 diabetes vs Healthy Control

[Edit](#) [Share](#) [Delete](#) [Restart](#)

Completed on March 5, 2024 in 27 minutes.

- Report**
- Input/Output
- Execution summary
- Genome Browser
- Api response

Sections

- Volcano plot
- Heatmap
- Diff expr genes
- PCA
- Correlation Plot
- GSEA GO
- GSEA Pathway

Volcano Plot

Filters

P field:

P-Value:

P-Value (-Log10): 1.30

Fold Change:

Down

Up

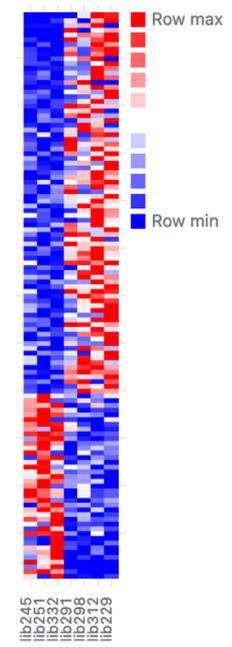
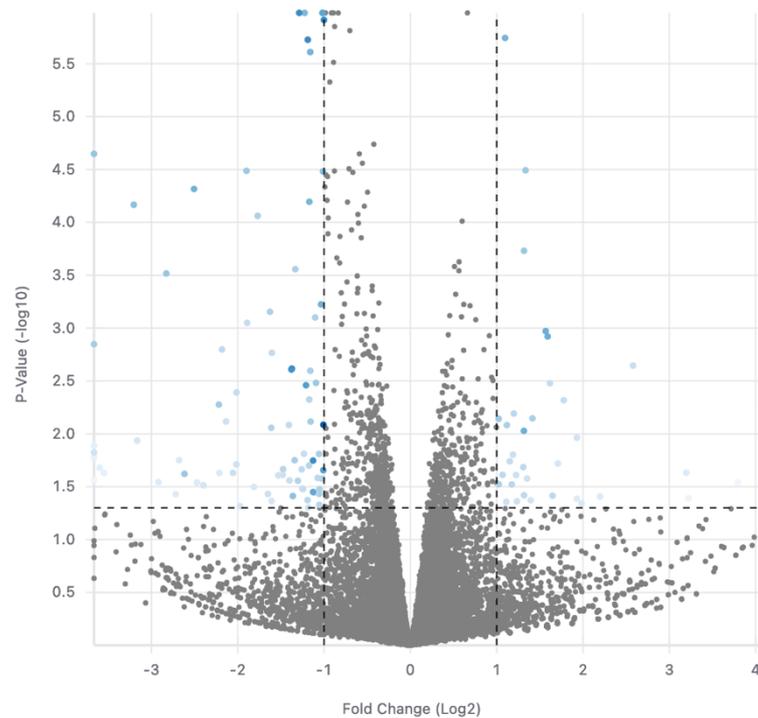
Fold Change (Log2): [-1.00 | 1.00]

Download up list

Download down list

Labels
Type gene symbols or ids to label and highlight them

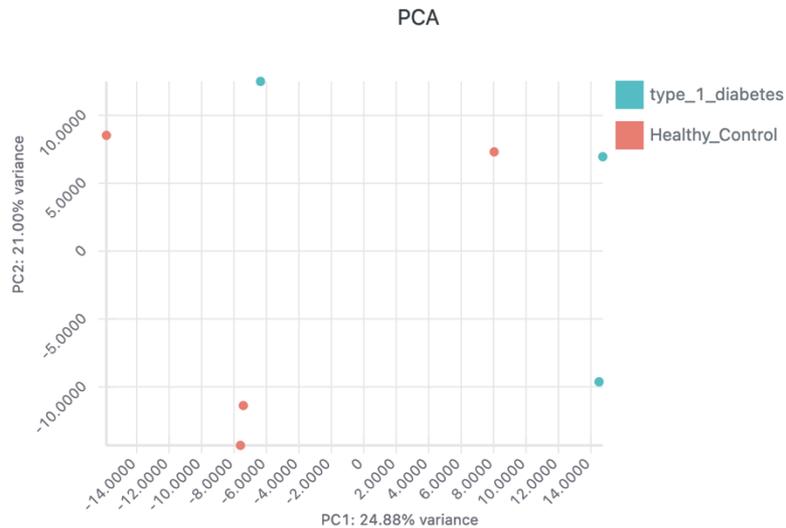
Total - 19180 / Up - 39 / Down - 80



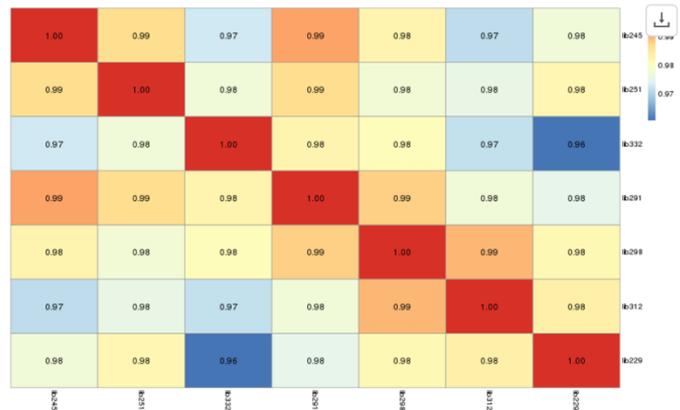
PCA axes

Select the axes you want to plot.

PC1 - PC2



Correlation Plot



type_1_diabetes					Healthy_Control			
Name	Q	Size	ES	NES	NOM p-val	FDR q-val	FWER p-val	Rank At Max
GOBP_MATURATION_O...		27	0.1268	0.4006	0.9494	0.9996	1.0000	14303
GOBP_MITOCHONDRIA...		157	0.0960	0.3135	0.9494	0.9998	1.0000	6210
HP_FOCAL_T2_HYPER...		32	0.1269	0.3613	0.8562	0.9999	1.0000	4029
GOMF_PROTEIN_DISU...		18	0.1452	0.4054	0.9470	0.9999	1.0000	13802
GOBP_RESPONSE_TO_...		21	0.5945	1.8093	0	1.0000	0.8520	1678
HP_STOMACH_CANCER		20	0.5324	1.7953	0	1.0000	0.8790	1710
GOBP_PREASSEMBLY_...		17	0.6547	1.7618	0	1.0000	0.9360	1437
GOBP_GPI_ANCHOR_M...		31	0.5002	1.7540	0	1.0000	0.9360	2303
GOMF_EXONUCLEASE_...		56	0.3759	1.7183	0	1.0000	1.0000	5746
GOMF_EXORIBONUCLE...		38	0.3536	1.7141	0	1.0000	1.0000	2726
GOBP_RESPONSE_TO_...		15	0.5706	1.6927	0	1.0000	1.0000	3051
GOBP_CYTOSKELETON...		79	0.3263	1.6869	0	1.0000	1.0000	2604
GOBP_MITOTIC_CYTO...		68	0.3407	1.6747	0	1.0000	1.0000	2604
GOCC_CAJAL_BODY		74	0.3755	1.6431	0	1.0000	1.0000	2760
HP_BREAST_CARCINOMA		59	0.3702	1.6337	0	1.0000	1.0000	1678
HP_ABNORMALITY_OF...		15	0.6320	1.6288	0	1.0000	1.0000	3414
HP_RHABDOMYOSARCOMA		21	0.4580	1.6273	0	1.0000	1.0000	2955
GOBP_CELLULAR_RES...		15	0.4852	1.6167	0.0515	1.0000	1.0000	4581
HP_APLASTIC_CLAVICLE		15	0.5400	1.6017	0.1333	1.0000	1.0000	2781
GOBP_BRANCHED_CHA...		22	0.5510	1.5974	0.0568	1.0000	1.0000	6141
GOMF_RNA_POLYME...		19	0.4472	1.5923	0	1.0000	1.0000	1752
HP_NEOPLASM_OF_TH...		78	0.3107	1.5799	0	1.0000	1.0000	3029
HP_ABNORMALITY_OF...		32	0.4929	1.5716	0	1.0000	1.0000	249
GOBP_CELLULAR_RES...		36	0.3768	1.5636	0	1.0000	1.0000	2006
GOBP_RNA_PHOSPHOD...		41	0.3527	1.5633	0	1.0000	1.0000	5746

GSEA Pathway

Download

[Export to CSV](#)

type_1_diabetes					Healthy_Control			
Name	Q	Size	ES	NES	NOM p-val	FDR q-val	FWER p-val	Rank At Max
REACTOME_SUMOYLAT...		44	0.4459	1.8516	0.0584	0.4868	0.3220	4813
REACTOME_SYNTHESI...		18	0.6531	1.7351	0	0.7765	0.6050	1437
REACTOME_G_BETA_G...		19	0.1666	0.4836	0.8848	0.9852	1.0000	4665
REACTOME_PROCESSI...		234	0.0976	0.4588	0.8475	0.9862	1.0000	7046
REACTOME_ADP_SIGN...		19	0.1682	0.4931	0.9339	0.9865	1.0000	4665
REACTOME_APC_C_CD...		73	0.1523	0.5282	0.8578	0.9894	1.0000	6266
REACTOME_NRF1_SIG...		15	0.1692	0.4942	0.9460	0.9902	1.0000	6139
REACTOME_HOST_INT...		125	0.0954	0.5017	0.9365	0.9917	1.0000	14556
REACTOME_RNA_POLY...		40	0.1040	0.5315	0.9410	0.9921	1.0000	6070
REACTOME_REGULATI...		67	0.1366	0.5088	0.8191	0.9929	1.0000	662
REACTOME_EPH_EPHR...		47	0.1646	0.5342	0.9244	0.9948	1.0000	5630
REACTOME_PROCESSI...		15	0.2200	0.5388	0.8337	0.9969	1.0000	15000
REACTOME_WNT5A_DE...		15	0.1570	0.5413	0.8668	0.9997	1.0000	6679
REACTOME_GO_AND_E...		26	0.6119	1.6447	0.0643	1.0000	0.8120	4440
REACTOME_HDR_THRO...		36	0.4844	1.6330	0	1.0000	0.8120	5415
REACTOME_TRANSCRI...		19	0.5634	1.5808	0.1255	1.0000	0.8710	4440
REACTOME_INTERACT...		34	0.3207	1.5598	0.1086	1.0000	1.0000	6502
REACTOME_BRANCHED...		21	0.5496	1.5302	0.1201	1.0000	1.0000	5725
REACTOME_G1_S_SPE...		28	0.5967	1.5259	0.0704	1.0000	1.0000	4440
REACTOME_APOPTOTI...		32	0.4033	1.5107	0.0638	1.0000	1.0000	1410
REACTOME_ACTIVATI...		37	0.5205	1.5060	0.0714	1.0000	1.0000	4359
REACTOME_TRANSCRI...		34	0.4299	1.4971	0.1250	1.0000	1.0000	7887
REACTOME_MEIOTIC_...		22	0.4973	1.4681	0	1.0000	1.0000	2977
REACTOME_APOPTOTI...		40	0.3761	1.4655	0.0680	1.0000	1.0000	2558
REACTOME_TRANSCRI...		16	0.5734	1.4369	0.0717	1.0000	1.0000	4440

Differential Expression (DESeq2, compare 2 groups) on type 1 diabetes vs Healthy Control

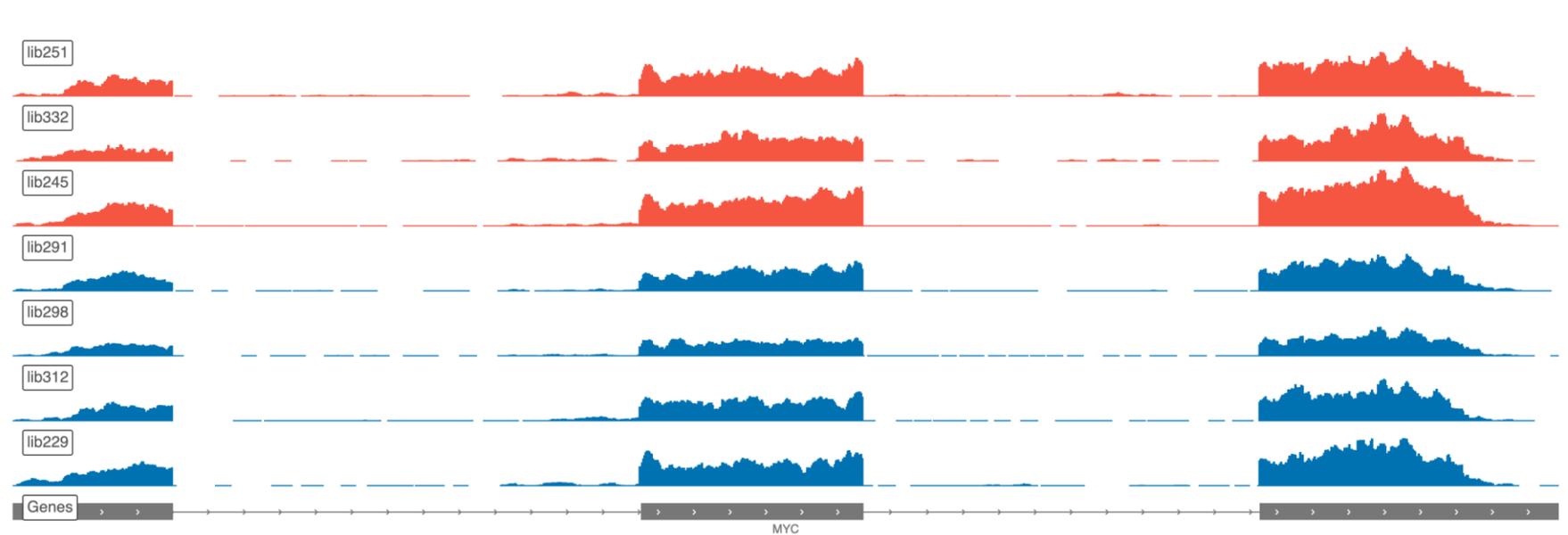
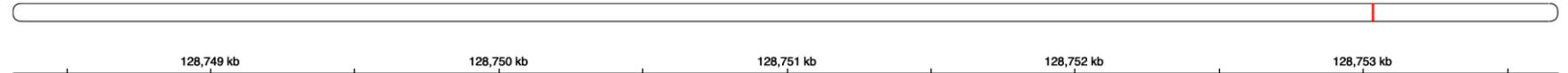
[Edit](#) [Share](#) [Delete](#) [Restart](#)

Completed on March 5, 2024 in 27 minutes.

Report Input/Output Execution summary **Genome Browser** Api response

[Equalize Scale](#) Add track sample ▾

IGV hg19 chr8 chr8:128,748,315-128,753,68 5,366 bp Cursor Guide Center Line Track Labels Save SVG ⊖ ⊕



Differential Expression (DESeq2, compare 2 groups) on type 1 diabetes vs Healthy Control

[Edit](#) [Share](#) [Delete](#) [Restart](#)

Completed on March 5, 2024 in 27 minutes.

[Report](#) [Input/Output](#) [Execution summary](#) [Genome Browser](#) [Api response](#)Pipeline [Differential Expression \(DESeq2, compare 2 groups\)](#)Projects [DRY解析教本_RNA-Seq](#)Samples [lib251](#)[lib332](#)[lib245](#)Controls [lib291](#)[lib298](#)[lib312](#)[lib229](#)

Files

 Show hidden

deseq

[deseq/Differential_Expression_DESeq2_compare_2_groups_on_type_1_diabetes_vs_Healthy_Control.cls](#)[deseq/corr_plot.png](#)[deseq/Differential_Expression_DESeq2_compare_2_groups_on_type_1_diabetes_vs_Healthy_Control.diffexpr.w_symbols.txt](#)[deseq/Differential_Expression_DESeq2_compare_2_groups_on_type_1_diabetes_vs_Healthy_Control.norm.gct](#)[deseq/Differential_Expression_DESeq2_compare_2_groups_on_type_1_diabetes_vs_Healthy_Control.pca-loadings.txt](#)[deseq/Differential_Expression_DESeq2_compare_2_groups_on_type_1_diabetes_vs_Healthy_Control.pca-rotation.txt](#)[deseq/Differential_Expression_DESeq2_compare_2_groups_on_type_1_diabetes_vs_Healthy_Control.vst.txt](#)

Create Lists

[lists/Diffexpr.min_count_10.pval_0.05.all_ids.txt](#)[lists/Diffexpr.min_count_10.pval_0.05.down.txt](#)[lists/Diffexpr.min_count_10.pval_0.05.up.txt](#)

GSEA

[gsea_go/Differential_Expression_DESeq2_compare_2_groups_on_type_1_diabetes_vs_Healthy_Control.GO.Gsea.zip](#)[gsea_go/Differential_Expression_DESeq2_compare_2_groups_on_type_1_diabetes_vs_Healthy_Control.pathway.gsea.zip](#)

GSEA (zip)



GSEA Report for Dataset filtered

Enrichment in phenotype: type_1_diabetes (3 samples)

- 1989 / 7514 gene sets are upregulated in phenotype **type_1_diabetes**
- 0 gene sets are significant at FDR < 25%
- 81 gene sets are significantly enriched at nominal pvalue < 1%
- 90 gene sets are significantly enriched at nominal pvalue < 5%
- [Snapshot of enrichment results](#) ← スナップショット
- Detailed [enrichment results in html](#) format ← リスト
- Detailed [enrichment results in TSV](#) format (tab delimited text)
- [Guide to](#) interpret results

Enrichment in phenotype: Healthy_Control (4 samples)

- 5525 / 7514 gene sets are upregulated in phenotype **Healthy_Contr**
- 0 gene sets are significantly enriched at FDR < 25%
- 302 gene sets are significantly enriched at nominal pvalue < 1%
- 694 gene sets are significantly enriched at nominal pvalue < 5%

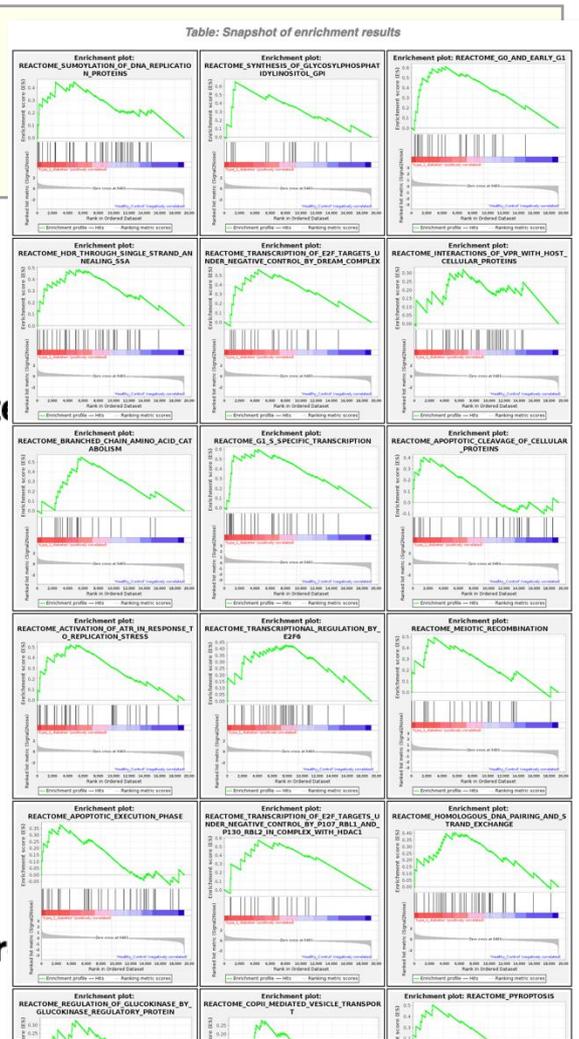
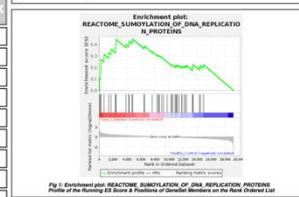


Table: Gene sets enriched in phenotype type_1_diabetes (3 samples) [plain text format]

RANK AT MAX	GS	follow link to MSigDB	GS DETAILS						
			SIZE	ES	NES	NOM p-val	FDR q-val	FWER p-val	RANK AT MAX
1	REACTOME_SUMOYLATION_OF_DNA_REPLICATION_PROTEINS		44	0.45	1.85	0.058	0.487	0.322	4813
2	REACTOME_SYNTHESIS_OF_GLYCOSYLPHOSPHATIDYLINOSITOL_GPI		18	0.65	1.74	0.000	0.777	0.605	1437
3	REACTOME_G0_AND_EARLY_G1		26	0.61	1.64	0.064	1.000	0.812	4440
4	REACTOME_HDR_THROUGH_SINGLE_STRAND_ANNEALING_SSA		36	0.48	1.63	0.000	1.000	0.812	5415
5	REACTOME_TRANSCRIPTION_OF_E2F_TARGETS_UNDER_NEGATIVE_CONTROL_BY_DREAM_COMPLEX		19	0.56	1.58	0.125	1.000	0.871	4440
6	REACTOME_INTERACTIONS_OF_VPR_WITH_HOST_CELLULAR_PROTEINS		34	0.32	1.56	0.109	1.000	1.000	6502
7	REACTOME_BRANCHED_CHAIN_AMINO_ACID_CATABOLISM		21	0.55	1.53	0.120	1.000	1.000	5725
8	REACTOME_G1_S_SPECIFIC_TRANSCRIPTION		28	0.60	1.53	0.070	1.000	1.000	4440
9	REACTOME_APOPTOTIC_CLEAVAGE_OF_CELLULAR_PROTEINS		32	0.40	1.51	0.064	1.000	1.000	1410
10	REACTOME_ACTIVATION_OF_ATR_IN_RESPONSE_TO_REPLICATION_STRESS		37	0.52	1.51	0.071	1.000	1.000	4359
11	REACTOME_TRANSCRIPTIONAL_REGULATION_BY_E2F6		34	0.43	1.50	0.125	1.000	1.000	7887
12	REACTOME_MEIOTIC_RECOMBINATION		22	0.50	1.47	0.000	1.000	1.000	2977
13	REACTOME_APOPTOTIC_EXECUTION_PHASE		40	0.38	1.47	0.068	1.000	1.000	2558
14	REACTOME_TRANSCRIPTION_OF_E2F_TARGETS_UNDER_NEGATIVE_CONTROL_BY_P107_RBL1_AND_P130_RBL2_IN_COMPLEX_WITH_HDAC1		16	0.57	1.44	0.072	1.000	1.000	4440
15	REACTOME_HOMOLOGOUS_DNA_PAIRING_AND_STRAND_EXCHANGE		41	0.40	1.43	0.178	1.000	1.000	5415
16	REACTOME_REGULATION_OF_GLUKOKINASE_BY_GLUKOKINASE_REGULATORY_PROTEIN		30	0.33	1.42	0.000	1.000	1.000	4727
17	REACTOME_COPII_MEDIATED_VESICLE_TRANSPORT		65	0.28	1.41	0.064	1.000	1.000	4430
18	REACTOME_PYROPTOSIS		23	0.51	1.41	0.107	1.000	1.000	2604
19	REACTOME_SHC_MEDIATED_CASCADE_FGFR4		15	0.53	1.39	0.039	1.000	1.000	3086
20	REACTOME_REGULATION_OF_TP53_ACTIVITY_THROUGH_METHYLATION		18	0.40	1.39	0.000	1.000	1.000	2955
21	REACTOME_RESOLUTION_OF_D_LOOP_STRUCTURES_THROUGH_SYNTHESIS_DEPENDENT_STRAND_ANNEALING_SDSA		25	0.42	1.37	0.128	1.000	1.000	4040
22	REACTOME_INTERACTIONS_OF_REV_WITH_HOST_CELLULAR_PROTEINS		35	0.27	1.36	0.212	1.000	1.000	4962
23	REACTOME_RESOLUTION_OF_D_LOOP_STRUCTURES		33	0.42	1.36	0.063	1.000	1.000	4040
24	REACTOME_ACTIVATION_OF_THE_PRE_REPLICATIVE_COMPLEX		33	0.50	1.35	0.129	1.000	1.000	4359
25	REACTOME_GLYOXYLATE_METABOLISM_AND_GLYCINE_DEGRADATION		25	0.42	1.34	0.162	1.000	1.000	5725
26	REACTOME_FRS_MEDIATED_FGFR3_SIGNALING		16	0.47	1.34	0.167	1.000	1.000	2883
27	REACTOME_CLASS_I_PEROXISOMAL_MEMBRANE_PROTEIN_IMPORT		20	0.39	1.34	0.097	1.000	1.000	981
28	REACTOME_NUCLEAR_IMPORT_OF_REV_PROTEIN		32	0.29	1.33	0.223	1.000	1.000	4727
29	REACTOME_FRS_MEDIATED_FGFR4_SIGNALING		17	0.50	1.33	0.041	1.000	1.000	3086
30	REACTOME_POLO_LIKE_KINASE_MEDIATED_EVENTS		16	0.54	1.31	0.203	1.000	1.000	3526
31	REACTOME_REGULATION_OF_TP53_ACTIVITY_THROUGH_PHOSPHORYLATION		88	0.28	1.30	0.134	1.000	1.000	3842
32	REACTOME_SUMOYLATION_OF_SUMOYLATION_PROTEINS		33	0.27	1.30	0.224	1.000	1.000	4727
33	REACTOME_INTERCONVERSION_OF_NUCLEOTIDE_DI_AND_TRIPHOSPHATES		29	0.38	1.29	0.174	1.000	1.000	3888
34	REACTOME_BUTYRATE_RESPONSE_FACTOR_1_BRF1_BINDS_AND_DESTABILIZES_MRNA		17	0.36	1.28	0.181	1.000	1.000	6202
35	REACTOME_E2F_MEDIATED_REGULATION_OF_DNA_REPLICATION		21	0.39	1.28	0.117	1.000	1.000	881
36	REACTOME_HYALURONAN_METABOLISM		16	0.47	1.28	0.185	1.000	1.000	3525
37	REACTOME_TRANSPORT_OF_VITAMINS_NUCLEOSIDES_AND_RELATED_MOLECULES		35	0.33	1.27	0.056	1.000	1.000	3163
38	REACTOME_PHASE_0_RAPID_DEPOLARISATION		23	0.40	1.27	0.058	1.000	1.000	3556
39	REACTOME_ERCC6_CS8_AND_EHMT2_G9A_POSITIVELY_REGULATE_RRNA_EXPRESSION		16	0.43	1.26	0.194	1.000	1.000	6159
40	REACTOME_CELL_CELL_JUNCTION_ORGANIZATION		40	0.37	1.25	0.104	1.000	1.000	1842
41	REACTOME_POSTMITOTIC_NUCLEAR_PORE_COMPLEX_NPC_REFORMATION		26	0.32	1.24	0.261	1.000	1.000	4727
42	REACTOME_GAMMA_CARBOXYLATION_HYPUSINE_FORMATION_AND_ARYLSULFATASE_ACTIVATION		37	0.31	1.24	0.078	1.000	1.000	3213
43	REACTOME_COBALAMIN_CBL_VITAMIN_B12_TRANSPORT_AND_METABOLISM		18	0.40	1.23	0.171	1.000	1.000	1197
44	REACTOME_FRS_MEDIATED_FGFR2_SIGNALING		18	0.43	1.22	0.199	1.000	1.000	2883
45	REACTOME_BASE_EXCISION_REPAIR_AP_SITE_FORMATION		17	0.37	1.22	0.205	1.000	1.000	4649

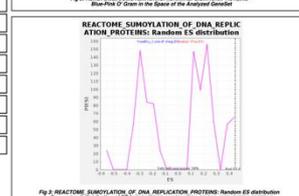
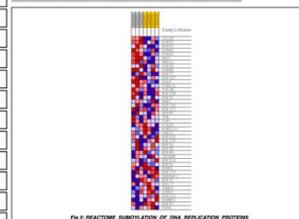
Gene Set Analysis Summary

Method	Gene Set Enrichment Analysis (GSEA)
Dataset	Phenotype: type_1_diabetes (3 samples)
Gene Set	REACTOME_SUMOYLATION_OF_DNA_REPLICATION_PROTEINS
Correlation	0.45
ES	0.45
ES (95% CI)	[0.32, 0.58]
ES (95% CI) (Lower)	0.32
ES (95% CI) (Upper)	0.58
ES (95% CI) (Mean)	0.45
ES (95% CI) (SD)	0.13
ES (95% CI) (SE)	0.02
ES (95% CI) (P)	0.000
ES (95% CI) (Q)	0.000
ES (95% CI) (R)	0.000
ES (95% CI) (S)	0.000
ES (95% CI) (T)	0.000
ES (95% CI) (U)	0.000
ES (95% CI) (V)	0.000
ES (95% CI) (W)	0.000
ES (95% CI) (X)	0.000
ES (95% CI) (Y)	0.000
ES (95% CI) (Z)	0.000



Gene Set Analysis Summary

Gene Set	ES	NES	NOM p-val	FDR q-val	FWER p-val	RANK AT MAX
REACTOME_SUMOYLATION_OF_DNA_REPLICATION_PROTEINS	0.45	1.85	0.058	0.487	0.322	4813
REACTOME_SYNTHESIS_OF_GLYCOSYLPHOSPHATIDYLINOSITOL_GPI	0.65	1.74	0.000	0.777	0.605	1437
REACTOME_G0_AND_EARLY_G1	0.61	1.64	0.064	1.000	0.812	4440
REACTOME_HDR_THROUGH_SINGLE_STRAND_ANNEALING_SSA	0.48	1.63	0.000	1.000	0.812	5415
REACTOME_TRANSCRIPTION_OF_E2F_TARGETS_UNDER_NEGATIVE_CONTROL_BY_DREAM_COMPLEX	0.56	1.58	0.125	1.000	0.871	4440
REACTOME_INTERACTIONS_OF_VPR_WITH_HOST_CELLULAR_PROTEINS	0.32	1.56	0.109	1.000	1.000	6502
REACTOME_BRANCHED_CHAIN_AMINO_ACID_CATABOLISM	0.55	1.53	0.120	1.000	1.000	5725
REACTOME_G1_S_SPECIFIC_TRANSCRIPTION	0.60	1.53	0.070	1.000	1.000	4440
REACTOME_APOPTOTIC_CLEAVAGE_OF_CELLULAR_PROTEINS	0.40	1.51	0.064	1.000	1.000	1410
REACTOME_ACTIVATION_OF_ATR_IN_RESPONSE_TO_REPLICATION_STRESS	0.52	1.51	0.071	1.000	1.000	4359
REACTOME_TRANSCRIPTIONAL_REGULATION_BY_E2F6	0.43	1.50	0.125	1.000	1.000	7887
REACTOME_MEIOTIC_RECOMBINATION	0.50	1.47	0.000	1.000	1.000	2977
REACTOME_APOPTOTIC_EXECUTION_PHASE	0.38	1.47	0.068	1.000	1.000	2558
REACTOME_TRANSCRIPTION_OF_E2F_TARGETS_UNDER_NEGATIVE_CONTROL_BY_P107_RBL1_AND_P130_RBL2_IN_COMPLEX_WITH_HDAC1	0.57	1.44	0.072	1.000	1.000	4440
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REACTOME_REGULATION_OF_GLUKOKINASE_BY_GLUKOKINASE_REGULATORY_PROTEIN	0.33	1.42	0.000	1.000	1.000	4727
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REACTOME_BASE_EXCISION_REPAIR_AP_SITE_FORMATION	0.37	1.22	0.205	1.000	1.000	4649



Expression count (STAR) on lib245

✎ Edit

🔗 Share

🗑 Delete

🔄 Restart

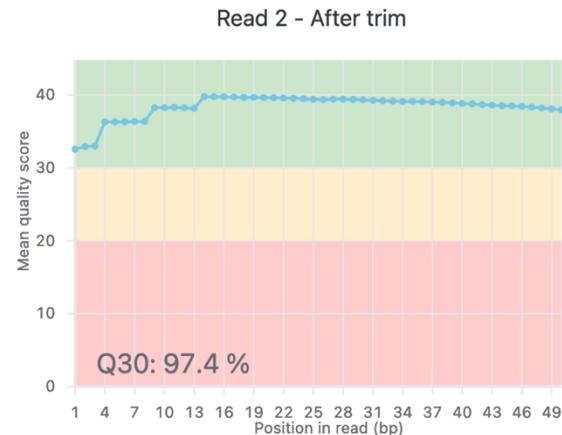
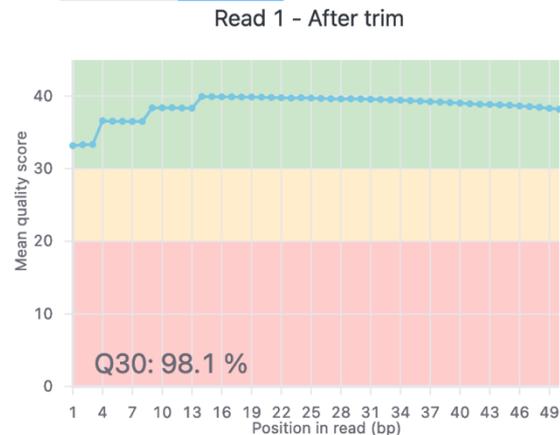
Completed on March 5, 2024 in 28 minutes.

[Report](#)[Input/Output](#)[Execution summary](#)[Api response](#)

Sections

[Quality scores](#)[Number of reads](#)[Metrics](#)[Read counts](#)[Genome browser](#)

Quality scores

Plot: Before trim After trim

This plot gives you a brief overview of sequence quality along all sequences for your sample. The y-axis in this plots, shows the quality scores.

The higher the score, the better the base call. The background of the graph divides the y-axis into very good quality scores (green), scores of reasonable quality (orange), and reads of poor quality (red).

The numbers in the y-axis represent the **probability of error**. A quality score of 10 means that there's a 10% chance that the base is incorrect. while 40 means that there's only a 0.01% probability of error. Remember. we are working with logs here.



- Quality scores

Number of reads

Metrics

Read counts

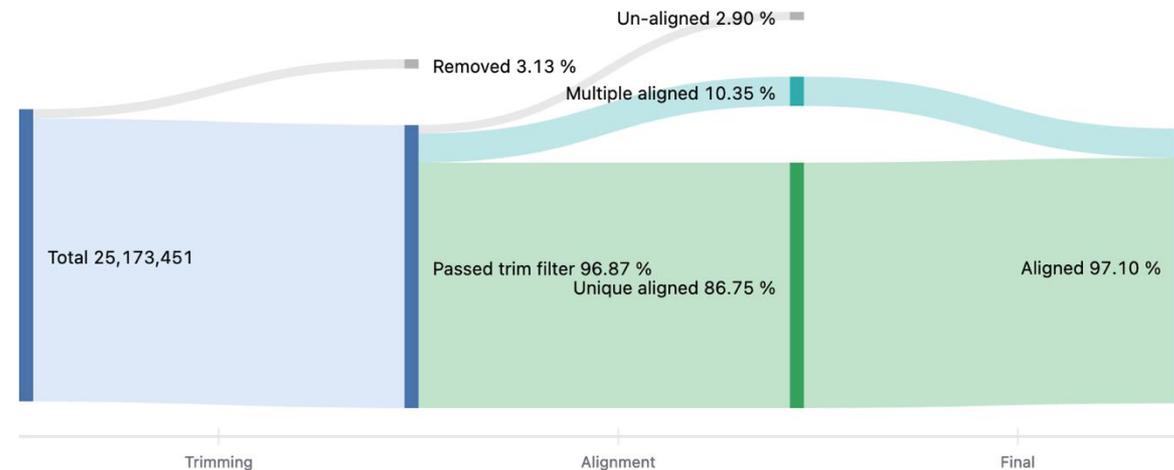
Genome browser

incorrect, while 40 means that there's only a 0.01% probability of error. Remember, we are working with logs here.

The sample here shows **96.88%** reads having quality score of 30 for R1 read & **95.48%** for R2 reads before filtering & reads after filtering **98.08%** reads having quality score of 30 for R1 reads & **97.36%** for R2 reads.

Here we observed the number of reads before filtering **50,346,902** & after quality filtering & trimming the number reads is **48,768,672**.

Number of reads



An important mapping quality parameter is the **percentage of mapped reads**, which is a global indicator of the overall sequencing accuracy and of the presence of contaminating DNA. We expect between 70% to 90% of regular RNA-seq reads to map onto reference genome, with a significant fraction of reads mapping to a limited number of identical regions equally well ('**multi-mapping reads**').

A total of **24,384,336** input reads were used for alignment, out of which, **21,154,149 (86%)** reads aligned uniquely.

The alignment seems to be good.

Metrics



Sections

Quality scores

Number of reads

Metrics

Read counts

Genome browser

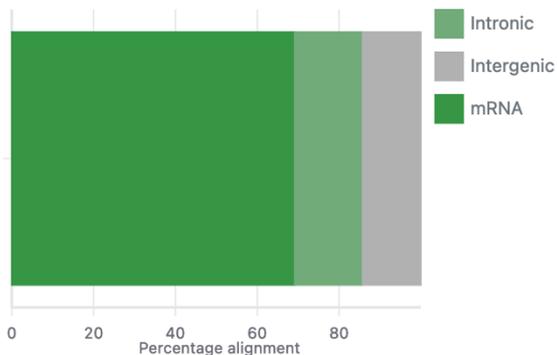
Trimming

Alignment

Final

An important mapping quality parameter is the **percentage of mapped reads**, which is a global indicator of the overall sequencing accuracy and of the presence of contaminating DNA. We expect between 70% to 90% of regular RNA-seq reads to map onto reference genome, with a significant fraction of reads mapping to a limited number of identical regions equally well (**'multi-mapping reads'**). A total of **24,384,336** input reads were used for alignment, out of which, **21,154,149 (86%)** reads aligned uniquely. The alignment seems to be good.

Metrics



This section reports how many alignments fall into **mRNA**, **intronic** and **intergenic regions**. Even if you have high genomic mapping rate for your sample, check to see where the reads are mapping.

Expect a high proportion of reads mapping to exonic regions (**> 60%**) and lower intronic mapping rates (**20 -30%**).

Here we observed **69.12%** reads aligned to coding (mRNA) region, **14.29%** to intergenic regions & **16.59%** to intronic regions.

Read counts

Count distribution: [By transcript](#) [By gene](#)

[Export to CSV](#)

Sections

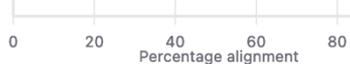
Quality scores

Number of reads

Metrics

Read counts

Genome browser



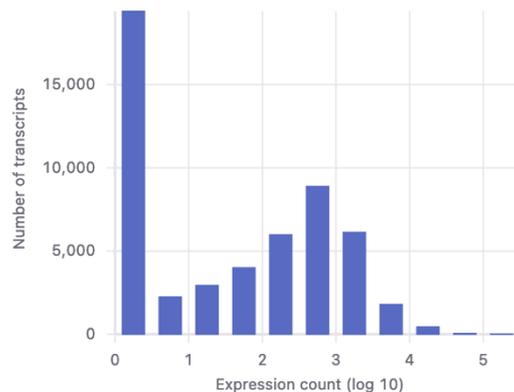
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Read counts

Count distribution: By transcript By gene



Export to CSV

Id	Symbol	Count	FPKM	TPM	
NM_001402	EEF1A1	178,665	1,410.36	3,149.87	view
NM_001286272	TPT1	105,763	609.08	1,360.30	view
NM_003295	TPT1	105,739	630.55	1,408.26	view
NM_001286273	TPT1	105,654	640.24	1,429.89	view
NM_004048	B2M	100,748	2,829.86	6,320.14	view
NM_000967	RPL3	67,304	1,394.54	3,114.53	view
NM_001033853	RPL3	66,579	1,549.79	3,461.25	view
NM_001961	EEF2	64,597	567.08	1,266.51	view
NR_046235	RNA45S5	63,310	131.40	293.47	view
NM_012423	RPL13A	61,363	1,441.69	3,219.82	view

Sections

- Quality scores
- Number of reads
- Metrics
- Read counts**
- Genome browser

Expression count (log 10)

NM_001033333	RPL3	66,373	1,943.73	3,461.23	view
NM_001961	EEF2	64,597	567.08	1,266.51	view
NR_046235	RNA45S5	63,310	131.40	293.47	view
NM_012423	RPL13A	61,363	1,441.69	3,219.82	view

< 1 2 3 4 5 ... 5178 >

Genome browser

[Equalize Scale](#) Add track sample ▾

IGV hg19 chr8 chr8:128,748,315-128,753,68 5,366 bp Cursor Guide Center Line Track Labels Save SVG - +

The visualization shows a genomic region on chromosome 8 (hg19) from 128,749 kb to 128,753 kb. The top track is labeled 'RNA-Seq Alignment' and shows signal peaks corresponding to gene expression. Below it is the 'Genes' track, which displays the structure of the MYC gene with arrows indicating the direction of transcription. A red vertical line is positioned at approximately 128,753,68 bp. On the right side, there are three gear icons for track settings.

所用時間比較

	自分で解析する※	Basepairを使う
所要時間	作業日数：9日 作業時間：13時間40分 サンプルファイル準備：約6日	ダウンロード：120分 STAR：平均24.8分 DESeq2：27分 実作業時間：10分
完了までの日数	30日	1日

※「次世代シーケンサーDRY解析教本」から算出

セットアップ（自分で解析する場合）

Mac Miniの場合	MacBook Pro 14inchの場合
<ul style="list-style-type: none">10コアCPU、16コアGPU、16コアNeural Engine搭載Apple M2 Proチップ32GBユニファイドメモリ1TB SSDストレージ	<ul style="list-style-type: none">12コアCPU、18コアGPU、16コアNeural Engine搭載Apple M3 Proチップ36GBユニファイドメモリ1TB SSDストレージ
274,800円（税込） キーボード、マウス、ディスプレイは別途	458,800円（税込）

※「次世代シークエンサーDRY解析教本」を参考に独自に選定

必要なパッケージ

- R：統計やグラフを描く
- Homebrew：パッケージ管理プログラム
- wget：ダウンロードするためのプログラム
- Bioconda：Pythonのパッケージ
- SRA Toolkit：SRAが提供するツール

API

他アプリケーションから操作

- サンプル、解析、結果
- アップロード、ダウンロード、実行、削除



```
import basepair

# CREATE SAMPLE
data = {
    "name": "Sample 10",
    "genome": "hg19",
    "datatype": "dna-seq",
    "file1": "/path/to/file1.fastq.gz",
    "file2": "/path/to/file2.fastq.gz",
}
sample_id = basepair.create_sample(data)

# RUN ANALYSIS
basepair.create_analysis(workflow_id=5, sample_id)
```

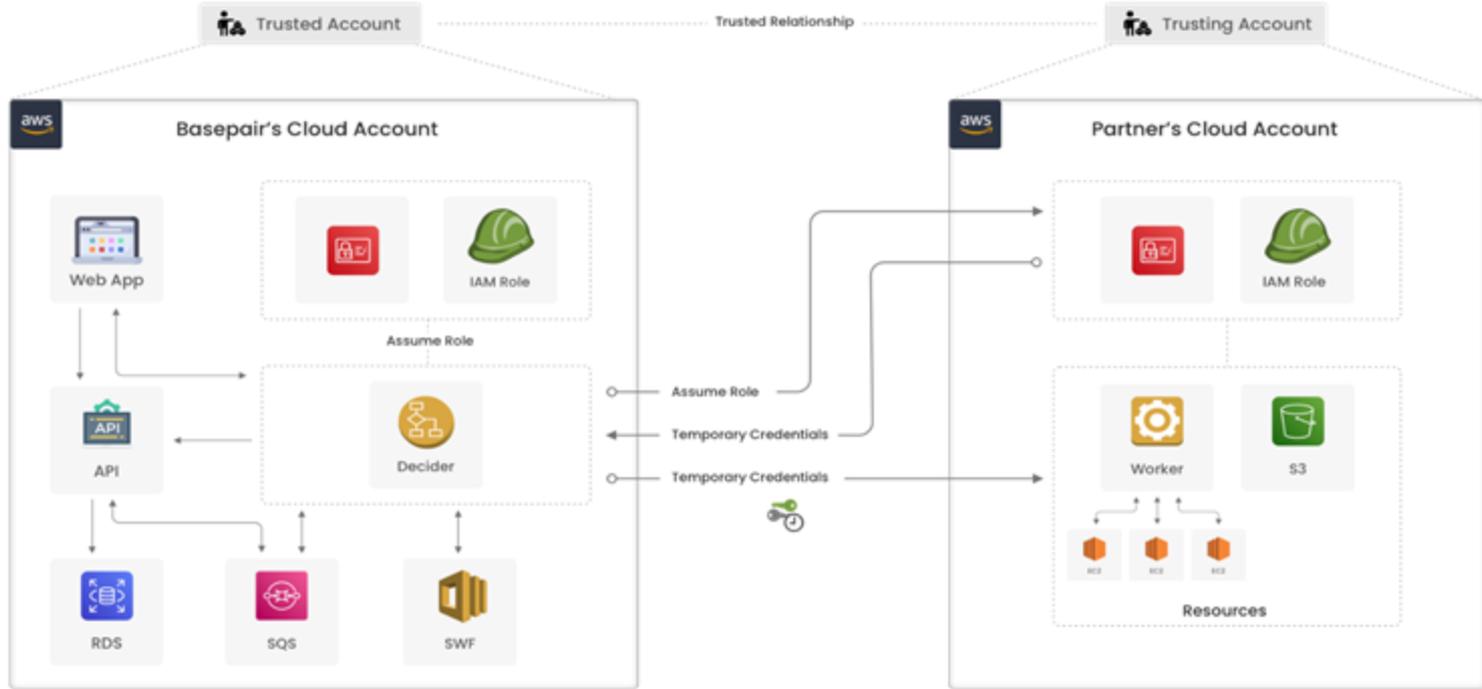
REST API

```
basepair --action create-sample --name "Sample 5" \
--datatype chip-seq --genome mm9 --file1
sample_5.1.fastq.gz

basepair --action create-analysis -w 14 -s 4111 -
s 4112 -c 5112 -c 5113 -c 5114
```

Python API

“Connected Cloud”



Basepairの利点

• セットアップ・インストール不要

- ファイルとインターネット環境があればOK

• 手元PCのスペックは不問

- サンプル数が多くても大丈夫
- クラウド上で計算が稼働するので、PCが固まったりしない
- 並列処理
- CLIでの操作も可能

• 共有可能

- チーム・共同研究相手に共有することができます



<https://basepairtech.jp/>